

---

# Magnitude of bacteraemia is associated with increased mortality in non-typhoid salmonellosis: a one-year follow-up study

---

KIM O. GRADEL,<sup>1,2</sup> CLAUS DETHLEFSEN,<sup>3</sup> HENRIK C. SCHØNHEYDER<sup>4</sup> and HENRIK NIELSEN<sup>1</sup>

Departments of <sup>1</sup>Infectious Diseases and <sup>4</sup>Clinical Microbiology, Aalborg Hospital, Aalborg, <sup>2</sup>Department of Clinical Epidemiology, Aarhus University Hospital, Aarhus, <sup>3</sup>Center for Cardiovascular Research, Aalborg Hospital, Aalborg, Denmark

Gradel KO, Dethlefsen C, Schønheyder HC, Nielsen H. Magnitude of bacteraemia is associated with increased mortality in non-typhoid salmonellosis: a one-year follow-up study. *APMIS* 2008;116:147–53.

We examined whether the number of positive bottles in a routinely used three-bottle blood culture (BC) set predicted one-year mortality in adult patients with non-typhoid *Salmonella* (NTS). Data from 1994 through 2003 in North Jutland County, Denmark, were retrieved from health databases and medical records. We used the number of positive BC bottles as an index of magnitude of NTS bacteraemia: Index 0 (reference) patients had a negative BC coincident with an NTS-positive faecal culture and index 1, 2, or 3 patients had increasing levels of NTS bacteraemia. For all patients and for patients with gastroenteritis we computed Kaplan-Meier curves to summarize survival over time and Cox regression analysis to estimate mortality in crude analyses and in analyses adjusted for comorbidity and age. There were 115, 43, 21, and 41 patients with index 0, 1, 2, and 3, respectively. One-year cumulative mortality was 4.4%, 14.0%, 28.6%, and 41.5% for indices 0 to 3. Adjusted one-year mortality rate ratios (with 95% confidence intervals) were 1.7 (0.5–5.8), 5.2 (1.5–17.4), and 5.3 (1.9–14.9) for index 1, 2, and 3 patients, respectively. These estimates remained robust for patients with gastroenteritis. We conclude that higher magnitude of bacteraemia predicted one-year mortality in NTS patients.

Key words: Magnitude of bacteraemia; non-typhoid salmonellosis; mortality.

Kim O. Gradel, Department of Clinical Epidemiology, Forskningens Hus, Aalborg Hospital, Aarhus University Hospital, Sdr. Skovvej 15, 9000 Aalborg, Denmark. e-mail: kog@rn.dk

Most studies focusing on predictors of outcome among patients hospitalized with non-typhoid *Salmonella* (NTS) have primarily addressed age, immunosuppression, and comorbid diseases (1). The complex interrelationship between age, immune status, comorbidity, and bacteraemia makes it difficult to separate the prognostic impact of each factor. Several older studies of various pathogenic bacteria reported that a higher magnitude of bacteraemia was associated with

increased mortality, but differences among groups were not taken into account, mortality after hospital discharge was not reported, and NTS bacteraemia was not addressed specifically (2–9). Recently we found that mortality rises as the number of positive blood culture (BC) bottles per initial BC set increases in patients with NTS bacteraemia (10). This led us to elaborate further on magnitude of NTS bacteraemia as a separate prognostic factor.

In the present study we made use of the local BC practice since 1992 in which 30 mL blood is obtained by venipuncture and then distributed

---

Received 28 June 2007.  
Accepted 18 November 2007.

equally between two aerobic bottles and one anaerobic bottle. Because *Salmonella* grows readily both aerobically and anaerobically, the number of positive bottles is an appropriate semi-quantitative index of bacterial density in the blood sample. Patients with a negative BC coincident with an NTS-positive faecal culture served as the reference cohort because of their confirmed NTS exposure and their physicians' attention to bacteraemia risk.

We examined whether magnitude of NTS bacteraemia was a prognostic factor for mortality during a one year follow-up period, taking into consideration comorbidity, age, and presence of gastroenteritis.

## METHODS

### *Study population*

The Danish health care system is tax financed and provides care free of charge for all residents. Acutely ill patients are admitted to the nearest hospital in their county of residence. During the 10-year study period (1994–2003), North Jutland County, Denmark, had an average population of ~500,000, served by eight public hospitals. Bacteriological services for the entire county were provided by the Department of Clinical Microbiology (DCM), Aalborg Hospital.

The bacteraemic cohort comprised all patients in North Jutland County with NTS isolated by BC during the study period (10). All were hospitalized in connection with their NTS bacteraemia. The reference cohort consisted of all inpatients in the County from 1995 through 2003 with NTS detected in a faecal culture and a negative BC drawn  $\pm 1$  day from the faecal culture. Patients with only one bottle sampled per BC were excluded, 24 without and 6 with NTS in the blood bottle, all of whom were 0–9 years old and alive after one year.

### *Record linkage*

All Danish residents have a unique 10-digit personal identification number (incorporating age and gender), which permits accurate linkage between relevant administrative and health registries. The Danish Civil Registration System contains continuously updated records of the vital status of all Danish residents, including date of emigration or death, if relevant, allowing us to ascertain patient status after one year of follow-up (11).

### *Microbiological procedures*

During the study period two BC systems were in use, the Colorbact® system (Statens Serum Institut [SSI], Copenhagen, Denmark) until December 1995 and the BacT/Alert® system (bioMérieux, Marcy l'E-

toile, France) thereafter (12). Both systems used two aerobic bottles and one anaerobic bottle per BC, except for younger children, for whom only one aerobic bottle was sampled (12, 13). In the Colorbact system the actual volumes sampled for the three bottles were 18–21 mL (12, 13), but after the introduction of the BacT/Alert system the target volume of 30 mL has been assured by regular quality control measurements (unpublished results).

Procedures for faecal cultures have been described in detail previously (14). In brief, faeces were cultured in NTS selective broth, followed by culture on indicative solid media and identification according to the Kaufmann-White scheme (15).

### *Microbiological data*

Identification of bacteraemic patients from the North Jutland County Bacteraemia Database has been described in detail previously (10, 16).

The national reference laboratory at SSI maintains a country-wide database with information on all NTS-positive cultures, including serotype and culture receipt date, but not culture type. After excluding 40 known NTS-bacteraemic patients, we assumed that all remaining cultures were faecal. Patients with NTS-positive faecal cultures were retrieved from this database from 1995 through 1997. For 1998–2003 we retrieved the same information on patients with NTS-positive faecal cultures from the DCM laboratory information system ADBakt (Autonik, Sködinge, Sweden). Unlike the SSI database, this database includes culture type. From 1995 on, all negative BC results were also recorded in the DCM laboratory information system.

### *Hospital discharge data*

Since 1977, non-psychiatric hospital discharge diagnoses have been recorded in the Hospital Discharge Registry of North Jutland, using the International Classification of Diseases (ICD) system [ICD-8 until 1994 and ICD-10 thereafter (ICD-9 was never implemented in Denmark)] (17). By linking patients with NTS-positive faecal cultures to this registry we were able to identify patients hospitalized with an NTS-related diagnosis  $\pm 1$  month of the culture receipt date. The main NTS-related diagnoses (NTS infection or diarrhoea due to presumed infectious cause) were retrieved using ICD-8 codes 003 and 009 and ICD-10 codes A02 and A09. Diagnoses of selected major comorbid diseases recorded in the 5-year period before the first NTS-positive culture were also retrieved for study patients. We classified comorbidity using the Charlson index, which includes 19 major disease categories (e.g., cardiovascular diseases, cancer, diabetes, and AIDS) and assigns scores to each of them (with higher scores associated with more severe disease categories) (18).

### *Medical records*

We used the medical records of the study patients to extract data on comorbidity, clinical and labora-

tory variables, and medicine administered during hospitalization (10).

#### Statistical analysis

The analytical unit was a patient, each of whom encountered the study population criteria only once. The initial event was receipt date of the first culture (BC for bacteraemic patients and faecal culture for reference cohort patients). The primary exposure was magnitude of bacteraemia as measured by the number of NTS-positive BC bottles (index 0, 1, 2, or 3) in the initial BC set. Outcome was death within 30, 180, and 365 days following the initial event.

Charlson index scores were categorized into three levels (0, 1–2, and >2 points) and four age groups were used (0–15, 16–64, 6–80, and >80 years).

We first estimated associations between the index and important covariates (age, gender, Charlson index scores, gastroenteritis, and main NTS serotype) using cross-tabulations. We then focused on Charlson index scores, age, and gastroenteritis, as these key covariates had been identified previously as prognostic factors for survival (10).

For the four index cohorts, we calculated crude cumulative mortality and cumulative mortality stan-

dardized to the distribution of Charlson index scores and age in the index 0 cohort.

We computed Kaplan-Meier survival curve estimates based on the index and key covariates. Cox proportional-hazards regression analysis was used to compute mortality rate ratios (MRR) and 95% confidence intervals (CI). First, the impact of the index was evaluated in crude analyses, after which we adjusted the model by including Charlson index scores and age. We repeated all analyses, excluding patients without gastroenteritis. Proportionality assumptions for all the models (assessed graphically and by Schoenfeld residuals) were found to be adequate (19).

Tests for trend for the indices 0–3, 0–2, and 1–3 were performed by repeating the adjusted Cox proportional-hazards regression analysis using the index as a continuous variable.

The software program Stata/SE 8.2 for Windows (Stata Corporation, College Station, TX, USA) was used for all analyses.

#### Ethical considerations

The study was conducted according to the guidelines of the regional scientific ethics committee for use of clinical and laboratory data, and approved by

TABLE 1. Main characteristics of the study population with non-typhoid Salmonella (NTS), North Jutland County, Denmark, 1994–2003

Factor	Index <sup>1</sup>				Total (%)
	0	1	2	3	
Age group (years)					
0–15	6 (5.2) <sup>2</sup>	1 (2.3)	0 (0)	2 (4.9)	9 (4.1)
16–64	79 (68.7)	23 (53.5)	11 (52.4)	13 (31.7)	126 (57.3)
65–80	23 (20.0)	12 (27.9)	8 (38.1)	15 (36.6)	58 (26.4)
>80	7 (6.1)	7 (16.3)	2 (9.5)	11 (26.8)	27 (12.3)
Gender					
Female	60 (52.2)	21 (48.8)	7 (33.3)	16 (39.0)	104 (47.3)
Male	55 (47.8)	22 (51.2)	14 (66.7)	25 (61.0)	116 (52.7)
Charlson index scores <sup>3</sup>					
0	81 (70.4)	23 (53.5)	10 (47.6)	15 (36.6)	129 (58.6)
1–2	29 (25.2)	14 (32.6)	7 (33.3)	19 (46.4)	69 (31.4)
>2	5 (4.4)	6 (14.0)	4 (19.1)	7 (17.1)	22 (10.0)
Gastroenteritis					
Yes	115 (100.0)	31 (72.1)	14 (66.7)	29 (70.7)	189 (85.9)
No	0 (0)	12 (27.9)	7 (33.3)	12 (29.3)	31 (14.1)
Main serotype					
<i>S. Enteritidis</i>	68 (59.1)	20 (46.5)	8 (38.1)	24 (58.5)	120 (54.6)
<i>S. Typhimurium</i>	36 (31.3)	4 (9.3)	4 (19.1)	8 (19.5)	52 (23.6)
Exotic <sup>4</sup>	11 (9.6)	19 (44.2)	9 (42.9)	9 (22.0)	48 (21.8)
Total no. patients	115	43	21	41	220

<sup>1</sup>Number of NTS-positive bottles in the initial blood culture drawn during hospital admission.

<sup>2</sup>Number of patients (%).

<sup>3</sup>Comorbidity as measured by Charlson index scores (18).

<sup>4</sup>All zoonotic serotypes other than *S. Enteritidis* and *S. Typhimurium*.

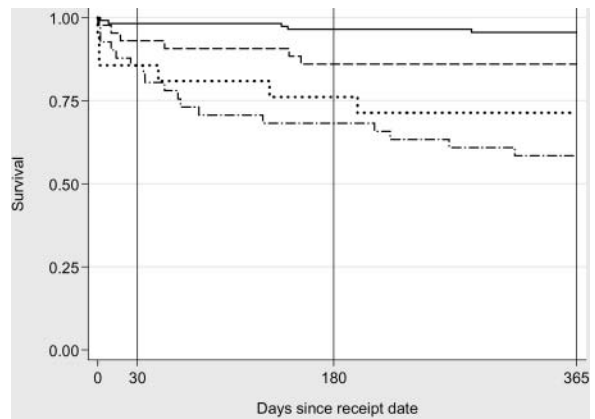
the Danish Data Protection Agency (Record no. 2004-41-4004).

## RESULTS

Among the study population of 220 patients, 115, 43, 21, and 41 had an index of 0, 1, 2, and 3, respectively (Table 1). Two index 1 patients had NTS detected in faeces 6 and 11 months after the actual NTS episode, but this did not result in hospital admission. Complete follow-up information was obtained for all patients except for one index 0 patient who emigrated on day 229.

The index rose with increasing age and level of Charlson index score (Table 1). While the study population had equal numbers of females and males, more males were represented among index 2 and index 3 patients. All index 0 patients, but only approximately 70% with indices 1 to 3, had gastroenteritis. *Salmonella* (*S.*) Typhimurium occurred more commonly than exotic serotypes (all serotypes other than *S. Enteritidis* or *S. Typhimurium*) in index 0 patients, while the reverse was seen in patients with indices 1 to 3. *S. Enteritidis* was equally distributed among the four index cohorts.

Regardless whether the follow-up period was 30, 180, or 365 days, cumulative mortality and MRR generally rose as the index increased (Table 2 & Fig. 1). The risk estimates remained



*Fig. 1.* Kaplan-Meier survival curves for 220 hospitalized patients with three bottles sampled for the initial blood culture drawn during hospital admission, North Jutland County, Denmark, 1994–2003. Index 0 (solid line): Patients with non-typhoid *Salmonella* (NTS) in a faecal culture and an NTS-negative blood culture drawn  $\pm 1$  day from the NTS-positive faecal culture. Indices 1 to 3: Patients with 1 (dashed line), 2 (dotted line), and 3 (dashed and dotted line) NTS-positive blood bottles, respectively. Receipt date is the date the faecal culture (index 0) or blood culture (indices 1 to 3) was received.

consistent in both the crude and adjusted analyses. In tests for trend, p-value for indices 0–3 was  $\leq 0.01$  after 180 and 365 days; this p-value remained the same in separate analyses for indices 0–2 and 1–3 after 365 days.

Because index 0 patients were selected according to an NTS-positive faecal culture only,

**TABLE 2.** Study population with non-typhoid *Salmonella*, North Jutland County, Denmark, 1994–2003: Cumulative mortality (CM), standardized cumulative mortality (SCM), mortality rate ratio (MRR) with 95% confidence intervals (CI) computed from Cox proportional-hazards regression analysis, and p for trend (for adjusted analyses)

Period	Index <sup>1</sup>	All patients					Gastroenteritis patients only				
		CM	SCM <sup>2</sup>	Crude MRR (95% CI)	Adjusted <sup>3</sup> MRR (95% CI)	p for trend	CM	SCM	Crude MRR (95% CI)	Adjusted MRR (95% CI)	p for trend
30-day	0	2/115 (1.7) <sup>4</sup>	1.7 <sup>5</sup>	1 (ref.)	1 (ref.)	0.07/	2/115 (1.7)	1.7	1 (ref.)	1 (ref.)	0.10/
	1	3/43 (7.0)	6.8	4.1 (0.7–24.5)	2.6 (0.4–16.0)	0.07/	1/31 (3.2)	6.5	1.8 (0.2–20.4)	1.4 (0.1–16.0)	0.30/
	2	3/21 (14.3)	7.0	9.1 (1.5–54.5)	5.5 (0.9–34.2)	0.54 <sup>6</sup>	1/14 (7.1)	10.4	4.3 (0.4–47.8)	2.4 (0.2–28.3)	0.32
	3	6/41 (14.6)	4.5	8.8 (1.8–43.4)	4.2 (0.8–22.3)		4/29 (13.8)	4.9	8.1 (1.5–44.4)	4.0 (0.7–23.2)	
180-day	0	4/115 (3.5)	3.5	1 (ref.)	1 (ref.)	0.01/	4/115 (3.5)	3.5	1 (ref.)	1 (ref.)	0.01/
	1	6/43 (14.0)	9.8	4.2 (1.2–14.9)	2.1 (0.6–7.7)	0.02/	3/31 (9.7)	9.1	2.8 (0.6–12.7)	1.9 (0.4–8.4)	0.23/
	2	5/21 (23.8)	13.5	7.8 (2.1–29.7)	4.8 (1.2–18.4)	0.16	2/14 (14.3)	11.7	4.4 (0.8–24.0)	2.2 (0.4–12.8)	0.12
	3	13/41 (31.7)	10.1	10.7 (3.5–32.8)	4.5 (1.4–14.4)		9/29 (31.0)	11.5	10.5 (3.2–34.1)	5.0 (1.5–16.9)	
365-day	0	5/114 (4.4) <sup>7</sup>	4.4	1 (ref.)	1 (ref.)	$<10^{-3}$ /	5/114 (4.4)	4.4	1 (ref.)	1 (ref.)	$10^{-3}$
	1	6/43 (14.0)	9.9	3.4 (1.0–11.1)	1.7 (0.5–5.8)	0.01/	3/31 (9.7)	9.2	2.3 (0.5–9.6)	1.5 (0.4–6.3)	0.08/
	2	6/21 (28.6)	20.2	7.8 (2.4–25.7)	5.2 (1.5–17.4)	0.02	3/14 (21.4)	18.4	5.4 (1.3–22.8)	3.2 (0.7–14.1)	0.01
	3	17/41 (41.5)	13.2	11.7 (4.3–31.6)	5.3 (1.9–14.9)		13/29 (44.8)	16.4	12.9 (4.6–36.3)	7.2 (2.5–20.6)	

<sup>1</sup> Blood culture density index, *cf* Table 1.

<sup>2</sup> Cumulative mortality standardized to distribution of age and Charlson index scores (18) in the index 0 cohort.

<sup>3</sup> Adjusted for Charlson index scores (18) and age.

<sup>4</sup> Number of deaths/total (%).

<sup>5</sup> %.

<sup>6</sup> Index 0–3/Index 0–2/Index 1–3.

<sup>7</sup> n=114, as one patient emigrated on day 229.

whereas index 1–3 patients were selected according to BC results, occurrence of gastroenteritis could confound the results. Of the 67 patients with indices 1 to 3 recorded after August 1997, 42/50 (84%) of those with gastroenteritis had faeces cultured, but only 2/17 (12%) of those without gastroenteritis had such cultures. When we repeated the analyses, excluding patients without gastroenteritis, the risk estimates were not affected (Fig. 2 & Table 2).

## DISCUSSION

Our study showed that higher magnitude of bacteraemia was associated with increased mortality among patients with NTS infections. These findings accord with bacteraemia studies in adults conducted several decades ago, which used very different methodologies (2–9). No study used multivariate analyses to control for potential confounders such as comorbidity and age, mortality after hospital discharge was not reported, and they did not include NTS among the diverse list of pathogens (*inter alia* *Staphylococcus aureus*, pneumococci, and Gram-negative rods) (2–9). In three studies, the prognostic impact of magnitude of bacteraemia in adults was reported for patient cohorts stratified by the authors' clinical evaluation of the prognosis (5–7). For Gram-negative bacteria, DuPont and Spink found that mortality increased in parallel with higher CFU/mL in patients with a good or

intermediate prognosis, while most patients with a poor prognosis died regardless of the magnitude of bacteraemia (5). Interestingly, when the same study group included all bacteraemic patients the same trend was reported for Gram-negative bacteria, but less clearly for Gram-positive bacteria or polybacteraemia (6).

Most studies focusing on magnitude of bacteraemia as a prognostic factor have relied on direct measurements of CFU/mL blood, derived either from pour-plate methods (2–7) or from plating on solid agar (20, 21). However, these quantitative methods are labour intensive and are rarely used for routine diagnostic purposes (22–24). Furthermore, their sensitivity is compromised by the small blood sample volumes (~1 mL). Because magnitude of bacteraemia in adults is low, culturing of 20–30 mL blood is recommended (23, 25–27).

Routine sampling of three blood bottles per BC in North Jutland County hospitals was implemented in 1992 (12, 13), spurred by American and Danish studies showing the critical importance of blood sample volume (25, 26). This approach has since been adopted by others (28). In our laboratory, all blood bottles were weighed from March 2000 through February 2002. Amongst the 1,173 weighed BC sets from this study there were no weight differences between BC indices 1, 2, and 3 (unpublished results). In addition, the index distribution did not differ when the two BC systems were compared (data not shown). We therefore believe this semi-quantitative index reflects bacterial density in the samples (26), and accordingly can be used as an appropriate measure of magnitude of NTS bacteraemia.

Some bacteraemia studies have attempted to relate cause of death to either infection or to factors such as underlying disease (29). This appears simplistic in view of the difficulty of separating the impact of individual factors on mortality among bacteraemic patients. The increasing cumulative mortality paralleling magnitude of NTS bacteraemia observed in our study up to one year after infection makes it difficult to argue that either microbial virulence or host response *per se* was responsible. Ours is one of the few studies to use a standard comorbidity index (the Charlson index) with proven value in prognostic studies (30). Still, we were not able to determine if magnitude of NTS bacteraemia was

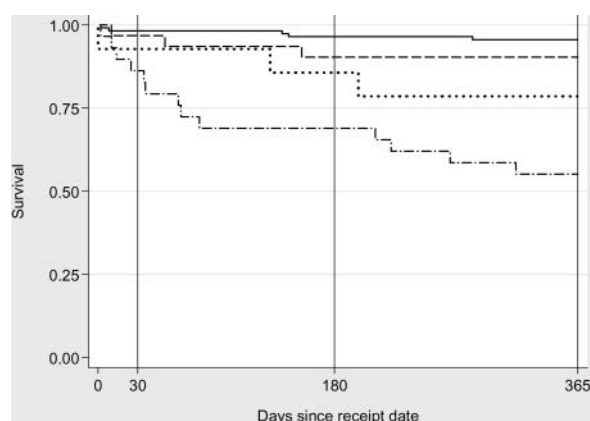


Fig. 2. Kaplan-Meier survival curves for 189 hospitalized gastroenteritis patients with three bottles sampled for the initial blood culture drawn during hospital admission, North Jutland County, Denmark, 1994–2003. Legend: see Fig. 1.

causally related to mortality or functioned as a marker of an underlying disease process unaccounted for in the Charlson index.

Several studies suggest that the absence of gastroenteritis worsens the prognosis for NTS bacteraemic patients (10, 31–34). However, when we restricted our analysis to gastroenteritis patients, the risk estimates remained essentially unchanged.

The main strengths of our study were the use of data from both administrative registries and medical records (which reduced possible misclassifications), virtually complete follow-up, and its applicability to the general population.

Our study also had a number of limitations. First, the wide CI reflect the limited precision of our risk estimates. Second, we also had to rely on previously identified confounders (10) without adjusting for other covariates. Third, as a higher proportion of NTS infections will probably remain undetected as their severity declines (35), the prevalence found in the reference cohort may be underreported in relation to the general population, though this is less important from the clinician's bedside point of view. Finally, while the definition of bacteraemic patients was clear-cut the novel approach used to identify the reference cohort may be questioned. We believe that narrowly limiting the time span between sampling of blood and faeces for culture and subsequently excluding patients without gastroenteritis made the reference cohort more homogeneous. However, this also resulted in exclusion of patients who with wider time spans might have had blood samples taken in connection with their clinical NTS symptoms.

## CONCLUSIONS

Increasing magnitude of NTS bacteraemia was associated with higher mortality after 30, 180, and 365 days. It is important to determine the prognostic utility of an analogous index derived from multiple BC sets with only two bottles each, which is used in several other hospitals and countries.

## REFERENCES

1. Hohmann EL. Nontyphoidal salmonellosis. *Clin Infect Dis* 2001;32:263–9.

2. Landsman JB. Bacteraemia and prognosis in lobar pneumonia: the results of quantitative blood culture in pneumococcus pneumonia. *Glasgow Med J* 1952;33:33–45.
3. Hall WH, Gold D. Shock associated with bacteremia; review of thirty-five cases. *AMA Arch Intern Med* 1955;96:403–12.
4. Weil MH, Spink WW. The shock syndrome associated with bacteremia due to gram-negative bacilli. *AMA Arch Intern Med* 1958;101:184–93.
5. DuPont HL, Spink WW. Infections due to gram-negative organisms: an analysis of 860 patients with bacteremia at the University of Minnesota Medical Center, 1958–1966. *Medicine (Baltimore)* 1969;48:307–32.
6. Kluge RM, DuPont HL. Factors affecting mortality of patients with bacteremia. *Surg Gynecol Obstet* 1973;137:267–9.
7. Kreger BE, Craven DE, Carling PC, McCabe WR. Gram-negative bacteremia. III. Reassessment of etiology, epidemiology and ecology in 612 patients. *Am J Med* 1980;68:332–43.
8. Whimbey E, Kiehn TE, Brannon P, Benezra D, Armstrong D. Clinical significance of colony counts in immunocompromised patients with *Staphylococcus aureus* bacteremia. *J Infect Dis* 1987;155:1328–30.
9. Schönheyder HC, Gottschau A, Friland A, Rosdahl VT. Mortality rate and magnitude of *Staphylococcus aureus* bacteremia as assessed by a semi-quantitative blood culture system. *Scand J Infect Dis* 1995;27:19–21.
10. Gradel KO, Schönheyder HC, Pedersen L, Thomsen RW, Nørgaard M, Nielsen H. Incidence and prognosis of non-typhoid *Salmonella* bacteraemia in Denmark: a 10-year county-based follow-up study. *Eur J Clin Microbiol Infect Dis* 2006;25:151–8.
11. Frank L. Epidemiology. When an entire country is a cohort. *Science* 2000;287:2398–9.
12. Pedersen G, Schönheyder HC, Kristensen B, Sørensen HT. Community-acquired bacteraemia and antibiotic resistance. Trends during a 17-year period in a Danish county. *Dan Med Bull* 2000;47:296–300.
13. Madsen KM, Schönheyder HC, Kristensen B, Sørensen HT. Secular trends in incidence and mortality of bacteraemia in a Danish county 1981–1994. *APMIS* 1999;107:346–52.
14. Gradel KO, Dethlefsen C, Schönheyder HC, Ejlersten T, Sørensen HT, Thomsen RW, et al. Severity of infection and seasonal variation of non-typhoid *Salmonella* occurrence in humans. *Epidemiol Infect* 2007;135:93–9.
15. Popoff MY, Le Minor L. Antigenic formulas of the *Salmonella* serovars. Institut Pasteur, Paris, France, 1997.
16. Schönheyder HC. [Two thousands seven hundred and thirty nine episodes of bacteremia in the

- county of Northern Jutland 1996–1998. Presentation of a regional clinical database]. *Ugeskr Laeger* 2000;162:2886–91.
17. Andersen TF, Madsen M, Jørgensen J, Mellemkjær L, Olsen JH. The Danish National Hospital Register. A valuable source of data for modern health sciences. *Dan Med Bull* 1999;46:263–8.
  18. Charlson ME, Pompei P, Ales KL, MacKenzie CR. A new method of classifying prognostic comorbidity in longitudinal studies: development and validation. *J Chronic Dis* 1987;40:373–83.
  19. Hosmer DW Jr, Lemeshow S. Assessment of model adequacy. In: Hosmer DW Jr, Lemeshow S, editors. *Applied Survival Analysis*. New York: John Wiley & Sons, Inc., 1999.
  20. Sullivan TD, LaScolea LJ Jr, Neter E. Relationship between the magnitude of bacteremia in children and the clinical disease. *Pediatrics* 1982;69:699–702.
  21. Sabui T, Tudehope DI, Tilse M. Clinical significance of quantitative blood cultures in newborn infants. *J Paediatr Child Health* 1999;35:578–81.
  22. Kiehn TE. Quantitative blood cultures. A review of 52 years. In: Brown AE, Armstrong D, editors. *Infectious Complications of Neoplastic Diseases: Controversies in Management*. New York: Yorke Medical Books, 1985.
  23. Yagupsky P, Nolte FS. Quantitative aspects of septicemia. *Clin Microbiol Rev* 1990;3:269–79.
  24. Lamy B, Roy P, Carret G, Flandrois JP, Delignette-Muller ML. What is the relevance of obtaining multiple blood samples for culture? A comprehensive model to optimize the strategy for diagnosing bacteremia. *Clin Infect Dis* 2002;35:842–50.
  25. Washington JA, Ilstrup DM. Blood cultures: issues and controversies. *Rev Infect Dis* 1986;8:792–802.
  26. Arpi M, Bentzon MW, Jensen J, Frederiksen W. Importance of blood volume cultured in the detection of bacteremia. *Eur J Clin Microbiol Infect Dis* 1989;8:838–42.
  27. Cockerill FRI, Wilson JW, Vetter EA, Goodman KM, Torgerson CA, Harmsen WS, et al. Optimal testing parameters for blood cultures. *Clin Infect Dis* 2004;38:1724–30.
  28. Arendrup M, Jensen IP, Justesen T. Diagnosing bacteremia at a Danish hospital using one early large blood volume for culture. *Scand J Infect Dis* 1996;28:609–14.
  29. Weinstein MP, Murphy JR, Reller LB, Lichtenstein KA. The clinical significance of positive blood cultures: a comprehensive analysis of 500 episodes of bacteremia and fungemia in adults. II. Clinical observations, with special reference to factors influencing prognosis. *Rev Infect Dis* 1983;5:54–70.
  30. de Groot V, Beckerman H, Lankhorst GJ, Bouter LM. How to measure comorbidity. a critical review of available methods. *J Clin Epidemiol* 2003;56:221–9.
  31. Ramos JM, Garcia-Corbeira P, Aguado JM, Arjona R, Ales JM, Soriano F. Clinical significance of primary vs. secondary bacteremia due to nontyphoid *Salmonella* in patients without AIDS. *Clin Infect Dis* 1994;19:777–80.
  32. Shimoni Z, Pitlik S, Leibovici L, Samra Z, Konigsberger H, Drucker M, et al. Nontyphoid *Salmonella* bacteremia: age-related differences in clinical presentation, bacteriology, and outcome. *Clin Infect Dis* 1999;28:822–7.
  33. Brown M, Eykyn SJ. Non-typhoidal *Salmonella* bacteraemia without gastroenteritis: a marker of underlying immunosuppression. Review of cases at St. Thomas' Hospital 1970–1999. *J Infect* 2000;41:256–9.
  34. Hsu RB, Tsay YG, Chen RJ, Chu SH. Risk factors for primary bacteremia and endovascular infection in patients without acquired immunodeficiency syndrome who have nontyphoid salmonellosis. *Clin Infect Dis* 2003;36:829–34.
  35. de Wit MA, Kortbeek LM, Koopmans MP, de Jager CJ, Wannet WJ, Bartelds AI, et al. A comparison of gastroenteritis in a general practice-based study and a community-based study. *Epidemiol Infect* 2001;127:389–97.